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| **Project title** | **Monitoring and managing insecticide resistance in UK pests** | | | |
| **Project number** | Cross sector: C&O 21510015; Potatoes 1120037; Horticulture 31120004 | | | |
| **Start date** | 1/4/12 | | **End date** | ongoing |
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| **Project aim and objectives** | | | | |
| The project is continuing to monitor the sensitivity of key pest species to insecticides to know which actives will work and which will not. The monitoring is being done primarily using bioassays on live samples. This approach is most effective because it provides an early indication of any reduced sensitivity to currently un-resisted insecticides in anticipation of the evolution of full-blown resistance that would lead to control failures. It is also independent of the need to know initially the exact mechanism of resistance.  Insect sampling has been done across GB through the continued involvement of stakeholders, including agronomy and agrochemical companies and sub-contractors. For some established resistance mechanisms, we are also continuing to use DNA-based diagnostics, which are specific for the mutations associated with particular resistance traits, and are incorporating any new such diagnostics as they become available (through other projects at Rothamsted). Samples of peach-potato aphids (*Myzus persicae*) are being screened for their response to cyantraniliprole, esfenvalerate, flonicamid, lambda-cyhalothrin, neonicotinoids, pymetrozine, spirotetramat and sulfoxaflor.  We are also screening other important aphid pests: potato aphids (*Macrosiphum euphorbiae*), currant-lettuce aphids (*Nasonovia ribisnigri*),willow-carrot aphids (*Cavariella aegopodii*), grain aphids (*Sitobion avenae*), bird cherry-oat aphids (*Rhopalosiphum padi*), and rose-grain aphids (*Metopolophium dirhodum*) when suspected insecticide control failures occur. Baseline bioassay data is being established for the relevant insecticides.  The project also now includes resistance monitoring in other important UK insect pests including cabbage stem flea beetles (*Psylliodes chrysocephala*), pea and bean weevils (*Sitona lineatus*), pollen beetles (*Meligethes aeneus*), diamond back moths (*Plutella xylostella*), silver Y moths (*Autographa gamma*) and onion thrips (*Thrips tabaci*).  The over-riding objective of the project is to retain the availability of effective pesticides by developing appropriate insect management strategies and providing robust scientific support to the regulatory decision making process. Guidance is regularly made available to advisors, growers and the scientific community through the [Insecticide Resistance Action Group (IRAG-UK)](https://ahdb.org.uk/knowledge-library/irag). Other routes of communication include articles in the trade press, presentations to growers and agronomists and papers in referred journals and conference proceedings (see below for this year’s outputs). More information on insecticide resistance is available from the [Insecticide Resistance Action Committee website.](https://www.irac-online.org/) | | | | |
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| **Key messages emerging from the project** | | | | |
| * Screening of peach-potato aphid (*M. persicae*) samples taken from the field and protected crops in 2018 showed that there continues to be no significant resistance (that may compromise control) to a range of compounds belonging to different chemical classes (cyantraniliprole, flonicamid, pymetrozine, spirotetramat and sulfoxaflor). Furthermore, there have been no significant shifts in response to diagnostic doses of these insecticides that are currently effective (un-resisted) in GB. * Strong pirimicarb resistance and pyrethroid resistance (conferred by MACE and super-kdr target site mechanisms respectively), remain prevalent in the *M. persicae* samples although there is evidence for some changes in the genetic make-up of the GB population with aphids carrying kdr alone becoming more common. * Microsatelitte analysis of *M. persicae* populations (an ‘in-kind’ contribution to the project) shows that the ‘O’ and ‘P’ super-clones seem to be on the wain with new genotypes, particularly ‘W’ (which also carries MACE and super-kdr), becoming prevalent. * Our findings continue to suggest that at least some *M. persicae* collected from protected crops may have come from more genetically-diverse, sexual populations on imported plant material. Obtaining samples from these environments remains very important as they are more likely to harbour aphids with new resistance mechanisms (e.g. to neonicotinoids) coming into the UK from abroad. * The baseline work on other important aphid pests continues to add data to the large database (which currently contains over 50 separate insecticide-susceptible baselines). These baselines will allow aphid pests linked to future reports of insecticide control problems to be quickly screened for potential resistance. * Greater pyrethroid resistance than that conferred by kdr has not been found in grain aphid (*S. avenae*) samples collected in 2018. * Pyrethroid resistance has been found in willow-carrot aphids (*C*. *aegopodii*). * Pyrethroid resistance continues to be seen in cabbage stem flea beetles (*P. chrysocephala*), pollen beetles (*M. aeneus*), pea and been weevils(*S. lineatus*) and diamond back moths (*P*. *xylostella*). * Pyrethroid resistance was not found in silver Y moth (*A. gamma*) samples. * Spinosad resistance, along with known pyrethroid resistance, was found for the first time in onion thrips (*T. tabaci*). This explains reports of reduced efficacy of this compound against this pest on salad onions and leeks. Growers, advisors and CRD have been made aware of these findings. | | | | |
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| **Summary of results from the reporting year** | | | | |
| * In 2018, we received, successfully reared and screened 28 open field and 5 protected crop peach-potato aphid (*M. persicae*)samples in England and Scotland (collected primarily by the sub-contractors and agronomy companies). The number of field samples was lower than in 2017 due to extreme climatic conditions (the ‘Beast from the East’ and the long hot, dry summer). * Screening bioassays applying diagnostic doses to live aphids from these samples continued to show no resistance to neonicotinoids, cyantraniliprole, flonicamid, pymetrozine, spirotetramat. Testing with sulfoxaflor, a compound added to the work in 2017, also showed no evidence of resistance. * In contrast, continued strong resistance to pirimicarb and pyrethroids was seen in most (> 70%) of the samples. * This was backed up by DNA tests showing that *M. persicae* carrying MACE resistance (to pirimicarb) and the new form (north European: *ne*) of super-kdr (conferring resistance to pyrethroids), with both mechanisms in the heterozygous form, continue to be common and widespread in the UK. * A few of the *M. persicae* field samples were found to contain aphids that were susceptible to lambda-cyhalothrin but resistant to esfenvalerate (both pyrethroid insecticides), with resistance specifically to esfenvalerate probably being caused by a new, as yet undisclosed, mechanism. * In the 2018 *M. persicae* field samples, there were a few (11%) *M. persicae* with extreme (R3) esterase-based resistance to organophosphates (OPs). 40% of the protected samples contained R3 aphids. * A comparison of the *M. persicae* insecticide resistance profiles found in the GB field versus protected crop samples showed that aphids with rarer combinations of resistance mechanisms/genotypes are found significantly more often at the protected sites. This is probably due to some of the aphids in these environments originating from more diverse, sexually-producing populations on imported plant material. * Micro-satellite testing (done in collaboration with the James Hutton Institute) showed that the *M. persicae* ‘O’ super-clone was no longer found in Scotland or England in 2018. This clone has been replaced by new super-clones; ‘S’ and ‘V’ (which carry kdr alone) and ‘W’ (which has the same resistance profile of MACE and super-kdr, both in the heterozygous form, as ‘O’ and ‘P’). The ‘P’ super-clone, which also has the same resistance genotype as ‘O’, has become rarer.The GB  *M. persicae* population appears, therefore, to be undergoing a change in its make-up. * *M. persicae* carrying strong (Nic-R++) neonicotinoid resistance, found in southern mainland Europe and north Africa, have so far not been seen in either the protected or field GB samples. However, there are recent unpublished reports of these aphids being found in Belgium. This highlights the importance of continued monitoring for these forms as they are strongly resistant to acetamiprid and thiacloprid sprays and to neonicotinoid seed treatments (the latter are available in protected environments). * We have continued to develop and validate the best bioassay method for various aphid species with the end product of insecticide-susceptible baselines for a large range of aphicides and aphid pests. These data will make quick screening bioassays available to assess whether any new reports of control failures against these aphid pests are due to the evolution of resistance. * Three grain aphid (*S. avenae*) samples (collected from winter wheat in Northamptonshire and Bedfordshire) in response to reports of pyrethroid control failures contained aphids with pyrethroid resistance but this remained at the expected level for aphids carrying kdr (in the heterozygous form). * No *S. avenae* kdr-RRs (homozygote) genotypes have been found to date. This may relate to a fitness cost associated with this genoype. * Four *C. aegopodii* samples, collected from Nottinghamshire, Suffolk and Yorkshire, contained aphids that were resistant to lambda-cyhalothrin applied at the 100% field rate. * Over forty cabbage stem flea beetle (*P. chrysocephala*)samples (collected from oilseed rape in England) were screened for pyrethroid resistance. The majority of these samples contained resistant adults. The hot spot of higher frequencies of resistant beetles in the south east of England was seen again with levels of resistance similar to 2017. * Two samples of pollen beetle (*M. aeneus*),collected from Essex and Leicestershire, contained adults carrying pyrethroid resistance. The response of these pollen beetles to lambda-cyhalothrin and tau-fluvalinate (pyrethroids) was similar. Bioassays applying a synergist prior to a pyrethroid gave good control demonstrating a metabolic-based mechanism is responsible for resistance in this pest. * Samples of striped flea beetle (*Phyllotreta striolata*), collected from Suffolk, turnip beetle (*Phyllotreta cruciferae*), collected from Suffolk, and three samples of bruchid beetle (*Bruchus rufimanus*), collected from Hertfordshire and Hampshire, were tested using lambda-cyhalothin. There is evidence for reduced sensitivity to the active in all three species but because of the small sample sizes used more testing is required before conclusions about pyrethroid resistance can be drawn. * Six diamond back moth (*P*. *xylostella*)samples (collected from England and Scotland) contained pyrethroid- resistant moths. However, there was no evidence of resistance to diamides and spinosad as there was 100% control in the bioassays with these two actives. * This resistance profile was the same as seen in  *P*. *xylostella* samples in 2016 and 2017 (also collected from England and Scotland) and supports reports from growers and agronomists that this pest is now overwintering in the UK, probably under netted brassicas. * Reports of reduced control of onion thrips (*T. tabaci*) with spinosad were followed up by tests on a sample (collected from salad onions in Worcestershire). Unlike in 2017, bioassays showed that none of the adults were resistant to either this insecticide or deltamethrin when applied at the recommended field rates. This may relate to the sample being collected further west than the previous samples * Two samples of silver Y moth (*Autographa gamma*), collected from Warwickshire in response to concerns of pyrethroid resistance, showed no evidence of resistance to lambda-cyhalothrin, spinosad or diamides. | | | | |
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| **Key issues to be addressed in the next year** | | | | |
| * Pymetrozine will be unavailable for use in the UK in the near future. As a result, monitoring with this compound will stop for the 2019 samples. * A one year project extension (with the same aims and approach) has been agreed with the funders. Three new partners (Certis, BASF and Procam) have joined the project consortium. Financial support is provided by all three AHDB crop sectors. | | | | |
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| **Lead partner** | | Rothamsted Research | | |
| **Scientific partners** | | Rothamsted Research | | |
| **Industry partners (for reporting year)** | | Adama, Agrii, AICC, AHDB-Cereals & Oilseeds, AHDB-Horticulture, AHDB-Potatoes, Bayer, BBRO, Belchim, Dow (Corteva), DuPont, Frontier, Hutchinsons, NuFarm, Sumitomo and Syngenta. | | |
| **Government sponsor** | | Chemicals Regulation Directorate/Defra (in-kind contribution). | | |

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| **Has your project featured in any of the following in the last year?** | |
| ***Events*** | ***Press articles*** |
| A Dewar The consequences of a total ban on neonicotinoid seed treatments for pest control in oilseed rape cereals and sugar beet in the UK *Syngenta Meeting,* Rougham, April 2019  R Harrington Monitoring of aphid populations as a basis for effective control. *IIRB (International Institute of Sugar Beet Research) Workshop:* *Growing sugar beet without neonicotinoid seed treatments*, Leuven, Belgium, March 2019  S White IPM of cabbage stem flea beetle. *Agtech Seminar*, Dunmow, March 2019  A Dewar The consequences of a total ban on neonicotinoid seed treatments for pest control in cereals and oilseed rape in the UK *Crop Management Partners*, Petersfield, March 2019   A Dewar The consequences of a total ban on neonicotinoid seed treatments for pest control in cereals and oilseed rapein the UK *Wessex Agronomy* Market Lavington, February 2019   A Dewar Consequences of the neonicotinoid ban in oilseed rape, cereals and sugar beet for pest control. *Boston and North Wash Training Group*, Old Leake, February 2019   S White Integrated pest management of cabbage stem flea beetle. *Syngenta iOSR Conference*, Northamptonshire, February 201.  L Field Monitoring and managing insecticide resistance in the UK. *Velcourt Annual Farmers Event*, Oxford, January 2019  S White: Managing OSR pests. *Farmers Weekly OSR Masters Live*.  Kensworth, December 2018  S Foster: Update on insecticide resistance in UK pests, including willow-carrot aphids. *British Carrot Growers Association R & D Committee Meeting*, Newark, November 2018  M Williamson & S Foster: Pyrethroid resistance in UK crop pests *Pyrethrum Meeting*, Cambridge, September 2018  S Foster: Pest control strategies, resistance issues, new actives. *Insect Pest Control Review Meeting*, NIAB, Cambridge, September 2018  S Cook: Ecologically-based Integrated Pest Management in oilseed rape: a need not an option! *Rothamsted Research Lecture*, Harpenden, June 2018  S Foster: Full ban on neonicotinoid seed treatments in open field crops: where do we go from here? *Bayer Stand at Cereals*, June 2018  M Stevens: *CIBE Technical Meeting*, Ghent, May 2018  S Foster: Monitoring and managing insecticide resistance in UK pests. *IRAG-UK Meeting*, Dunmow, April 2018  R Collier: Aphid control in leafy salads: trials in SCEPTREplus. *British Leafy Salad Association - Annual General Meeting*, Stoneleigh, April 2018 | Cabbage stem flea beetles, *NIAB-TAG Agronomy Update*,September 2018  Virus looms in OSR, *Crop Production Magazine*, September 2018  Farming without neonicotinoid seed treatments, *Bayer Crop Focus Magazine*, August 2018  How will the neonic seed treatment ban affect beet? *Farmers Weekly*, June 2018  Tips to manage BYDV in cereals without neonics, *Farmers Weekly*, June 2018  Overcoming resistance in a post-neonic world, *Bayer Website*, June 2018  Resistance is far from futile, *The Grower Magazine*, June 2018  New insights into insecticide resistance, *Crop Production Magazine*, April 2018 |
| ***Conference presentations, papers or posters*** | ***Scientific papers*** |
| S Foster Winter oilseed rape without neonicotinoids. *NORBARAG*, Malmo, Sweden, March 2019.  S Foster Control of the cabbage flea beetles and peach-potato aphids in the UK: development of insecticide resistance and alternative methods to control these pests. *Danish Crop Production Congress,* Denmark, January 2019  S Foster Update on insecticide resistance in UK pests. *AICC Conference*, Towcester, January 2019  S Foster: Organiser of Session and introductory talk at AAB Meeting: Pest control after the loss of the neonicotinoid seed dressings. *Crop Protection in Southern Britain*, Brighton, November 2018  S Foster: Insecticide resistance in UK pests: the good, the bad and the ugly *European Congress of Entomology*, Naples, July 2018  M Stevens: *IIRB Summer Congress*, France, June 2018 | LE Walsh, MT Gaffney, GL Malloch, **SP Foster**, MS Williamson & G Purvis. First evidence of retained sexual capacity and survival in the pyrethroid resistant *Sitobion avenae* (F.) (Hemiptera: Aphididae) SA3 super-clone following exposure to a pyrethroid at current field-rate. *Irish Journal of Agricultural and Food Research. In Press* |
| ***Other*** | |
| **Guidelines and Alerts**  IRAG-UK: Insecticide resistance status in UK potato crops, *Guideline*  IRAG-UK: Insecticide resistance status in UK cereal crops, *Guideline*  IRAG-UK: Insecticide resistance status in UK oilseed rape crops, *Guideline*  IRAG-UK: Insecticide resistance and its management, *Guideline*  Rothamsted News Release: Thrips know their onions, *Alert*  AHDB News: Pyrethroid resistance in willow-carrot aphid, *Alert*  AHDB News: Spinosad resistance found in onion thrips, *Alert*  AHDB SCEPTREplus Blog: Targeting onion thrips, *Alert*  **Radio**  *Farming Today*: *BBC Radio 4*: Pyrethroid resistance in Cabbage Stem Flea Beetles, September 2018 | |